



PRODUCTION OF RECOMBINANT ENZYMES IN BACTERIAL HOSTS

Università degli Studi di Urbino
23 maggio 2024



Our Main recombinant enzymes

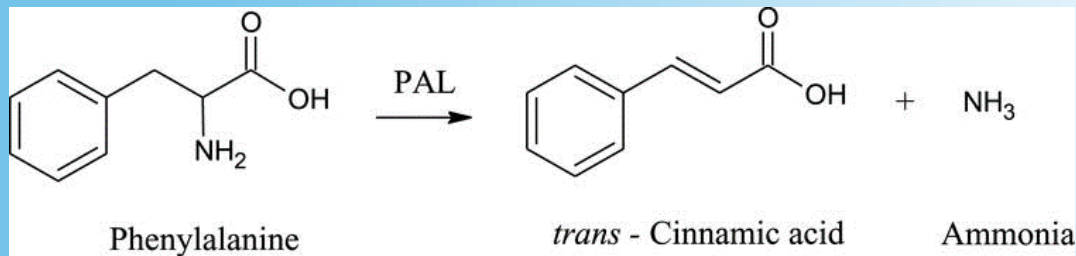
- **PAL: Phenylalanine Ammonia Lyase**
- **GAMT: GuanidinoAcetate MethylTransferase**
- **MAT2A: Methionine AdenosylTransferase 2A**
- **EDCas9: Enhanced Deletion Cas9**

All are expressed in *Escherichia coli* BL21(DE3)

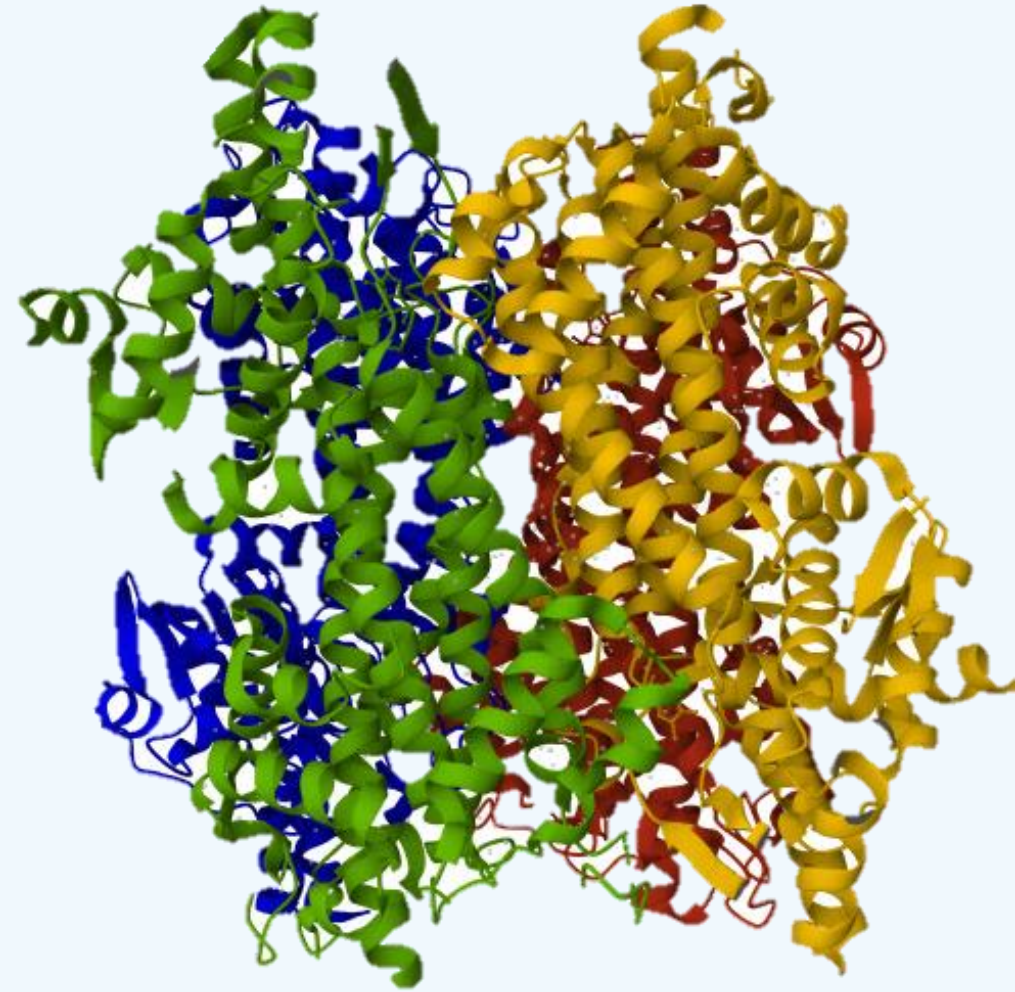
PAL

Phenylalanine Ammonia Lyase

- sequence derived from the cyanobacterium *Anabaena variabilis*
- converts L-Phenylalanine into *trans*-cinnamic acid and ammonia



- potential treatment for Phenylketonuria

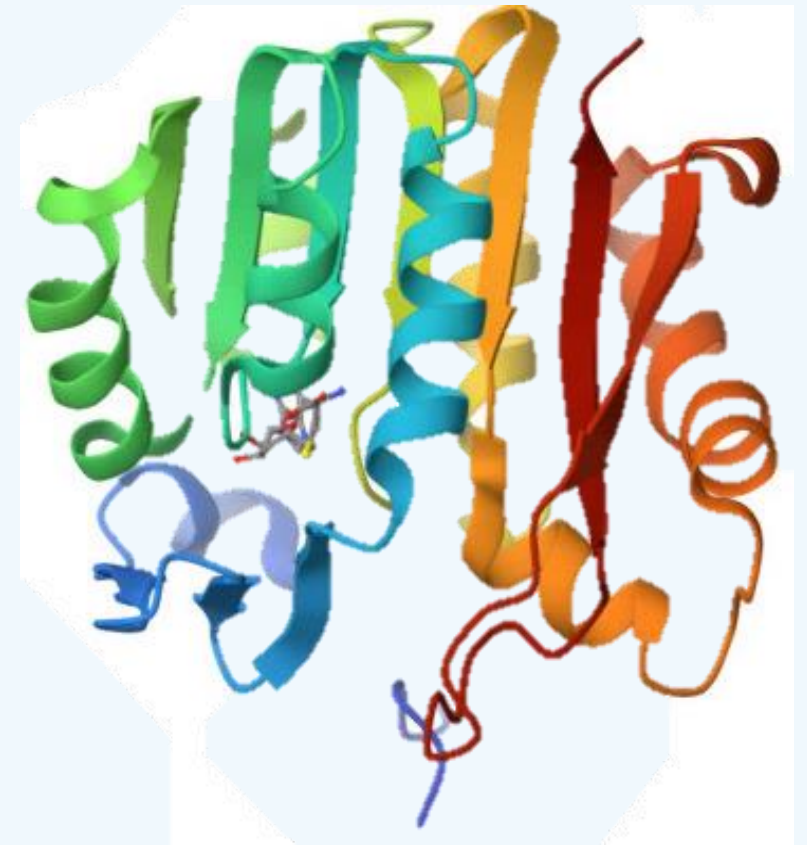
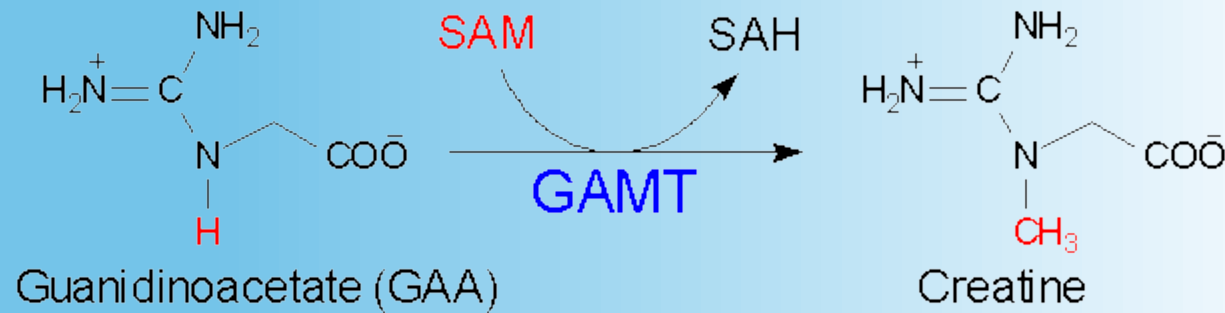


GAMT

GuanidinoAcetate MethylTransferase

- originally derived from the human gene, 4 aminoacids have been mutated to increase stability and solubility

- converts Guanidinoacetate and S-Adenosyl-Methionine into Creatine and S-Adenosyl-Homocysteine

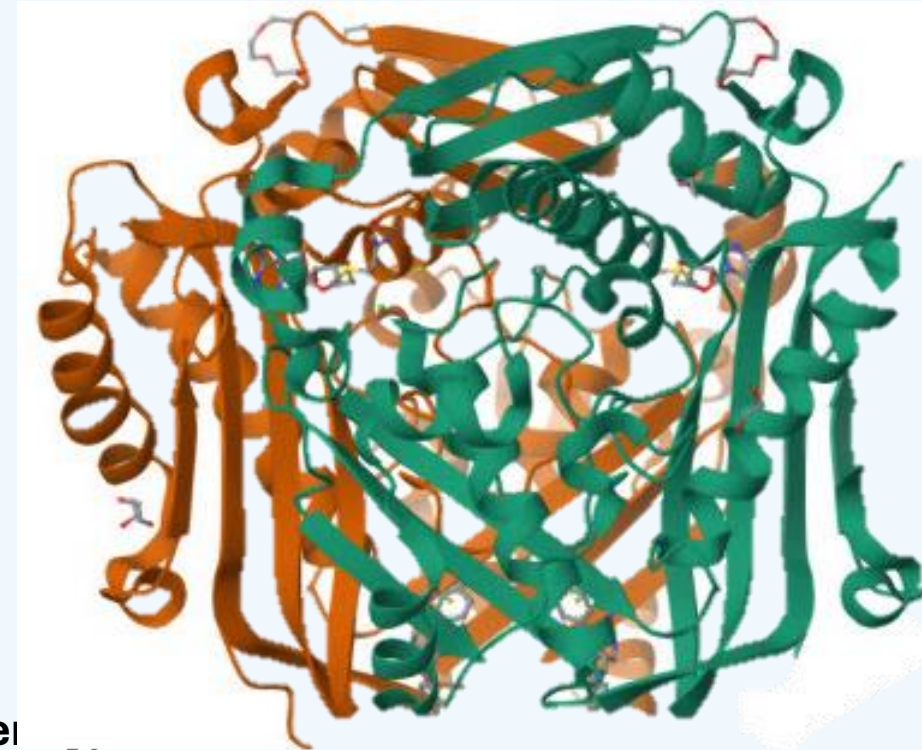
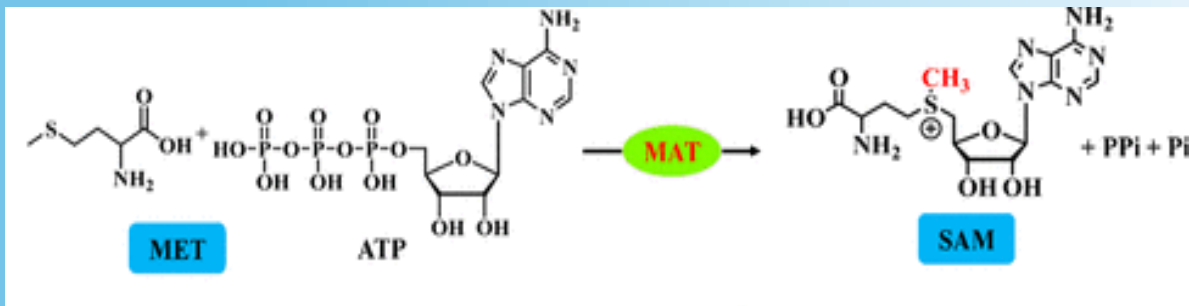


- to be administered in conjunction with MAT2A

MAT2A

Methionine AdenosylTransferase 2A

- derived from the human MAT2A sequence, with no modifications
- Forms S-Adenosyl-Methionine from Methionine and ATP.



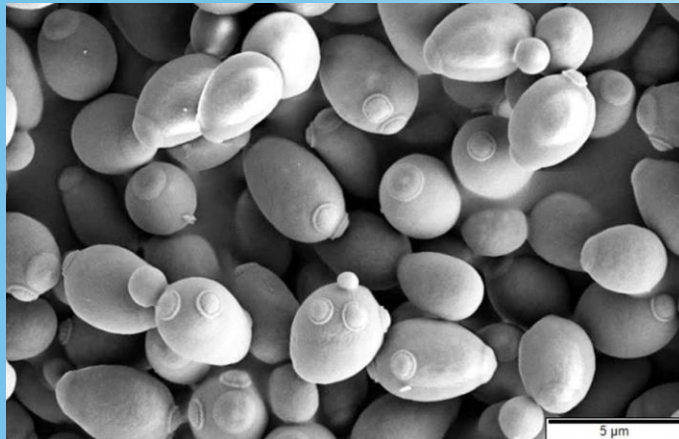
- to be used in conjunction with GAMT as a therapy for GAMT-deficiency. MAT2A supplies SAM, which GAMT needs to produce creatine



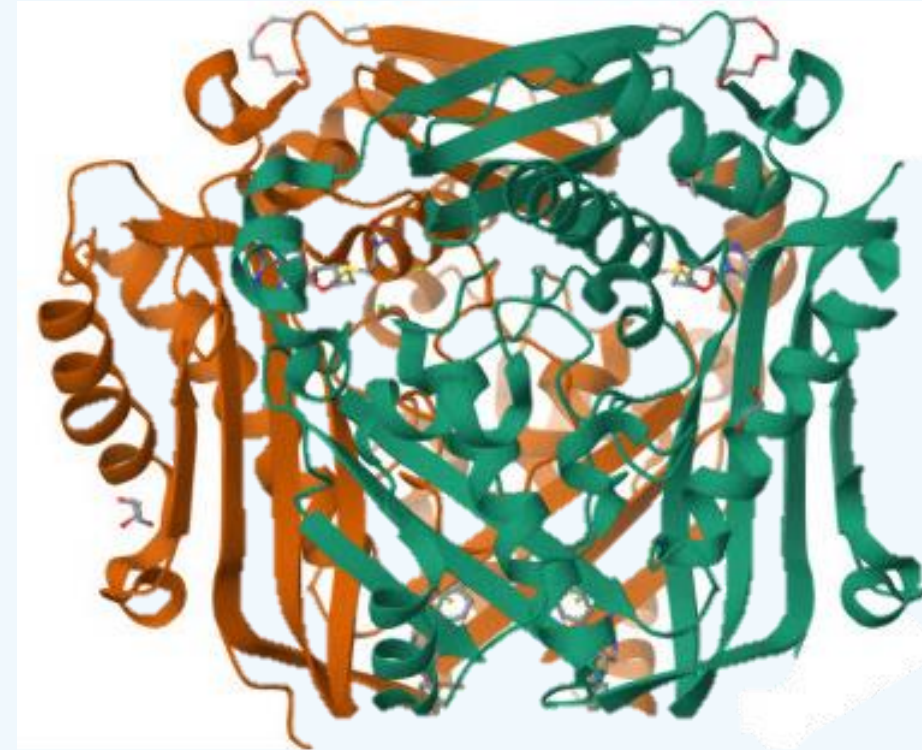
As MAT2A is inactive when expressed in bacteria, production has been moved to *Saccaromyces cerevisiae*

Advantages over Escherichia Coli:

- no contamination from bacterial Endotoxins
- capable of post-translational modifications
- cost-effective and safe



Thanks to:
Dott. Gianluca Morganti
Dott.ssa Federica Biancucci
Prof. Michele Menotta

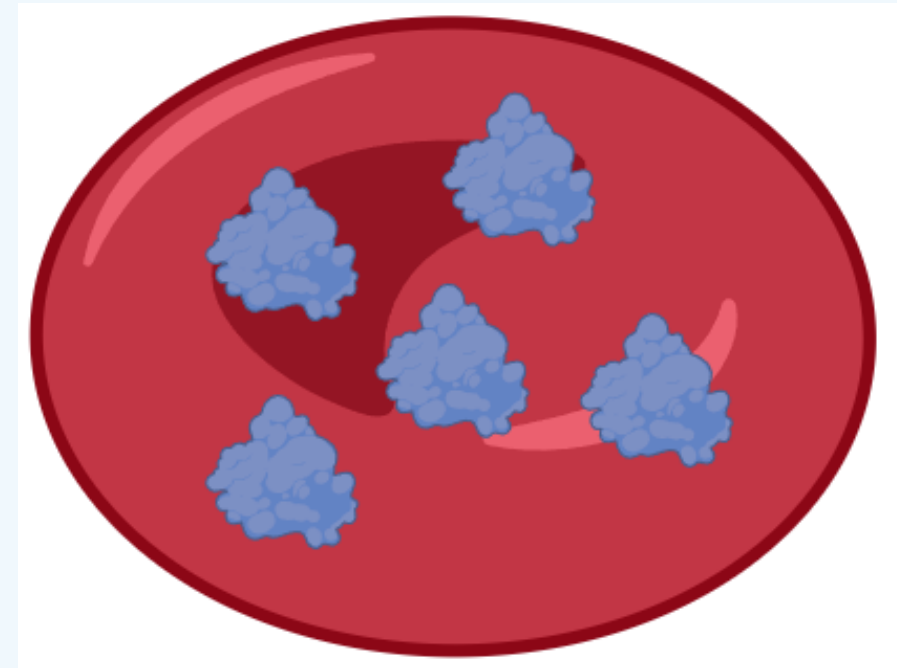


Erythrocytes as preferred delivery method

Enzyme loading into erythrocytes,
which are later reinfused into the
patient

Advantages:

- erythrocytes provide a “safe” environment
- longer half-life, which translates into fewer administrations
- fewer adverse affects, such as immune activation



EDCas9

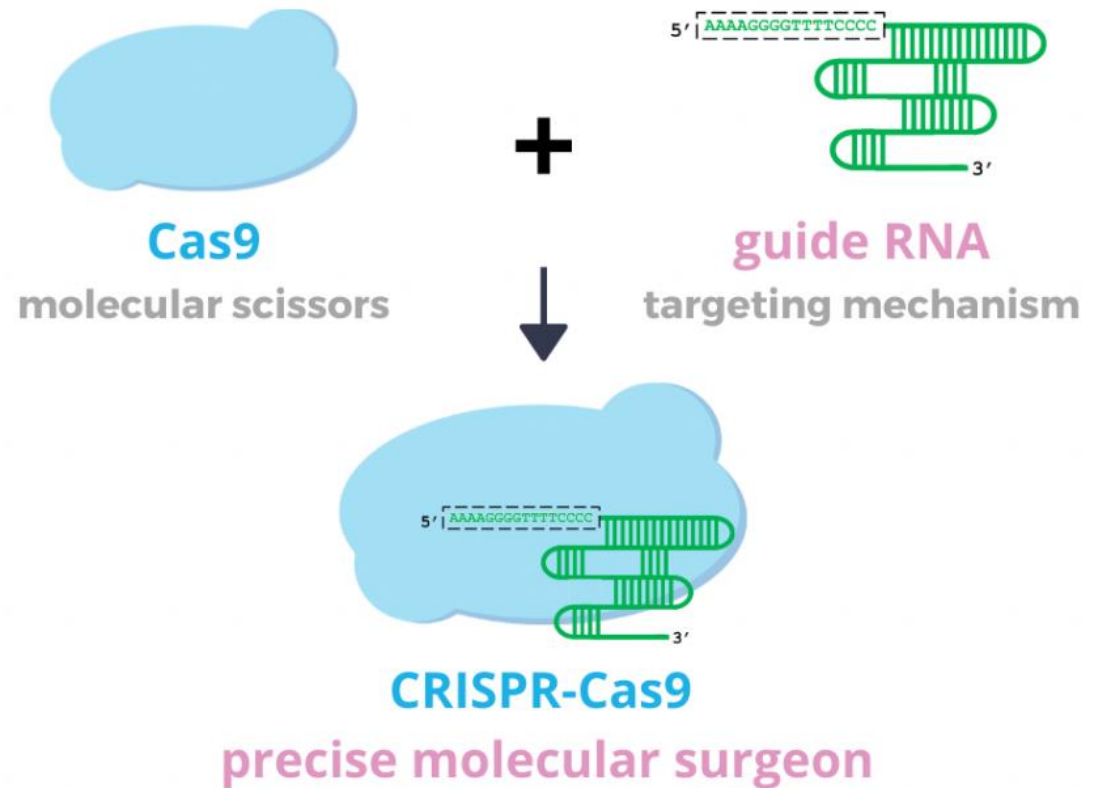
Enhanced Deletion Cas9

- Derived from the acclaimed CRISPR/Cas9 genome editing system
- fusion between Cas9 endonuclease and TREX2 endonuclease
- when coupled with a specific gRNA, the ribocomplex targets a pathologic variant of the USH2A gene, thus correcting the mutation



Thanks to:
Pietro de Angeli, Ph.D.

Components in CRISPR-Cas9



Recombinant protein production workflow

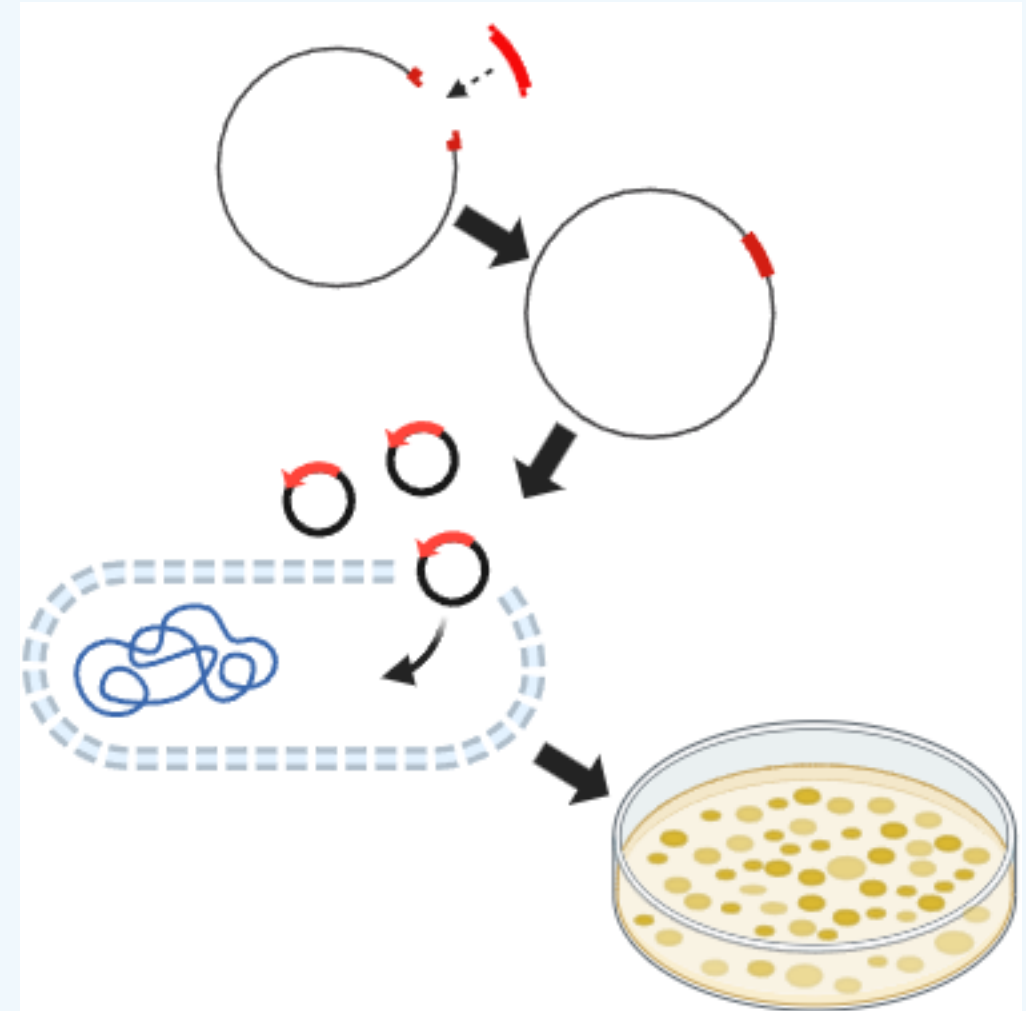
Protein-coding sequence is inserted into a plasmid



The plasmid is transformed into competent bacterial cells



Successfully transformed cells are selected in agar plates



Recombinant protein production workflow: Microbial culture processing

Transformed cells are cultured in a flask or bioreactor

Protein expression is induced through chemicals, such as IPTG

After a set amount of time, cells are harvested, lysed and centrifuged

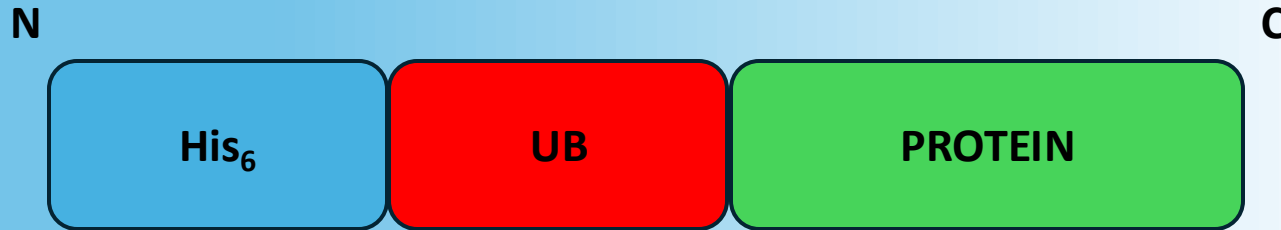
The clarified lysate is loaded onto a FPLC (Fast Protein Liquid Chromatography) column



Recombinant protein production workflow: protein purification

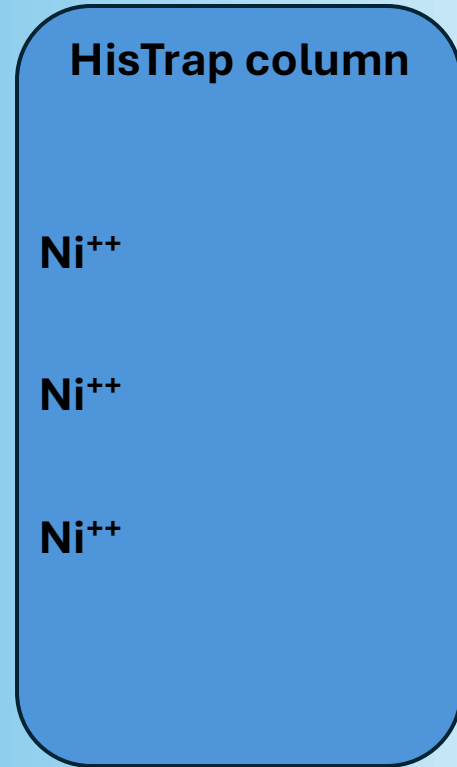
Most of our recombinant proteins are expressed with a **His-Ubiquitin tag** as a fusion partner

The Histidine portion of the tag allows purification of the recombinant protein through IMAC



IMAC: Immobilized Metal Affinity Chromatography

Cell lysate =
His-Ub-Protein + contaminants



The cell lysate is loaded onto the HisTrap column

IMAC: Immobilized Metal Affinity Chromatography



His-trap column

Ni⁺⁺ His-Ub-Protein

Ni⁺⁺ His-Ub-Protein

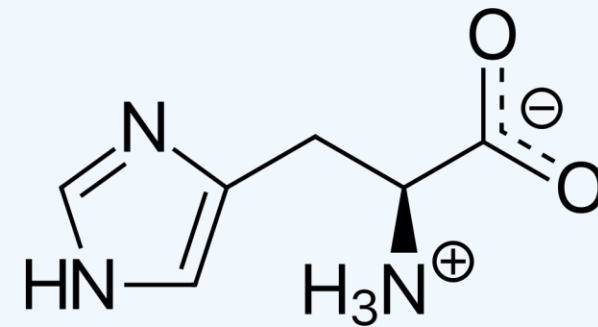
Ni⁺⁺ His-Ub-Protein



Contaminants

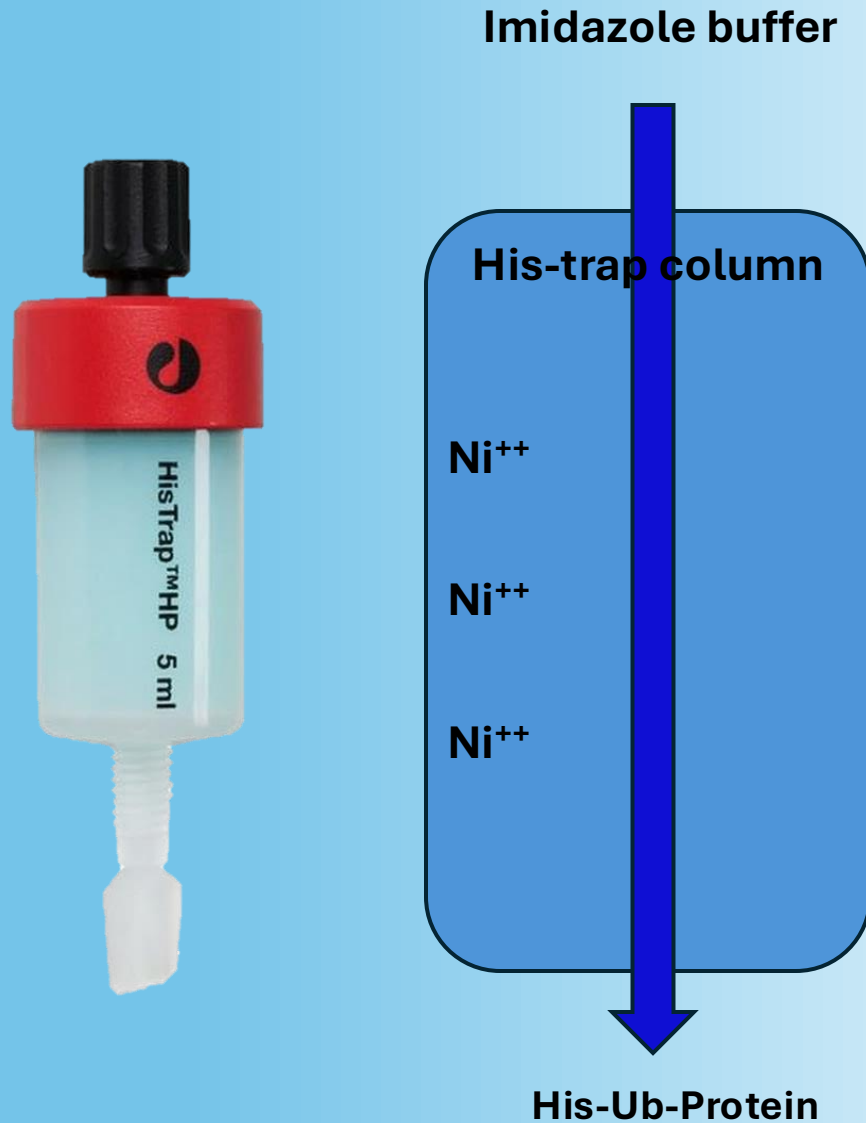
**The histidine portion of the recombinant protein
binds the nickel ions attached to the column
resin**

The contaminants are eliminated

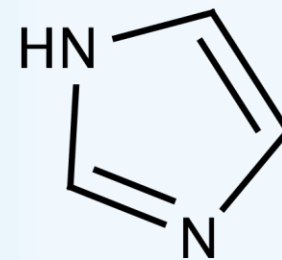


Histidine

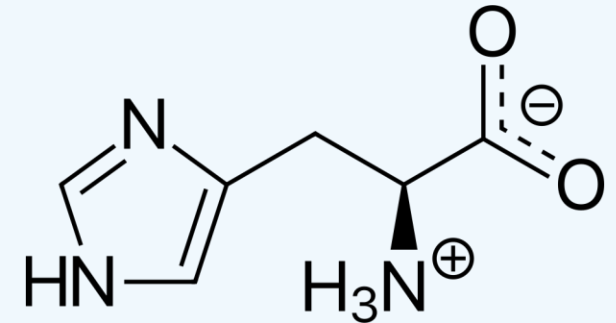
IMAC: Immobilized Metal Affinity Chromatography



The recombinant protein is detached from the resin through an imidazole buffer

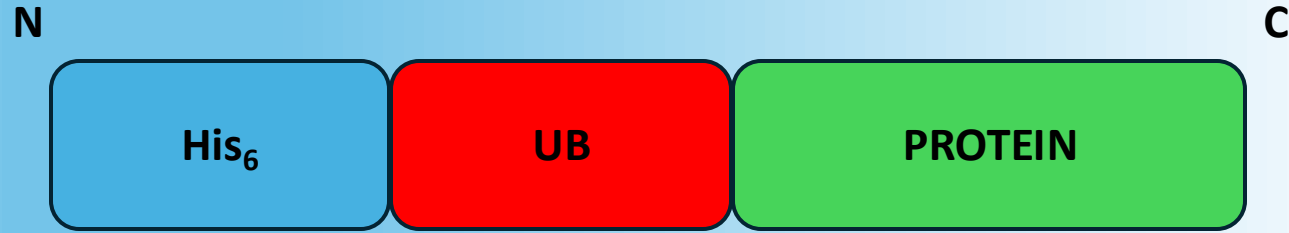


Imidazole

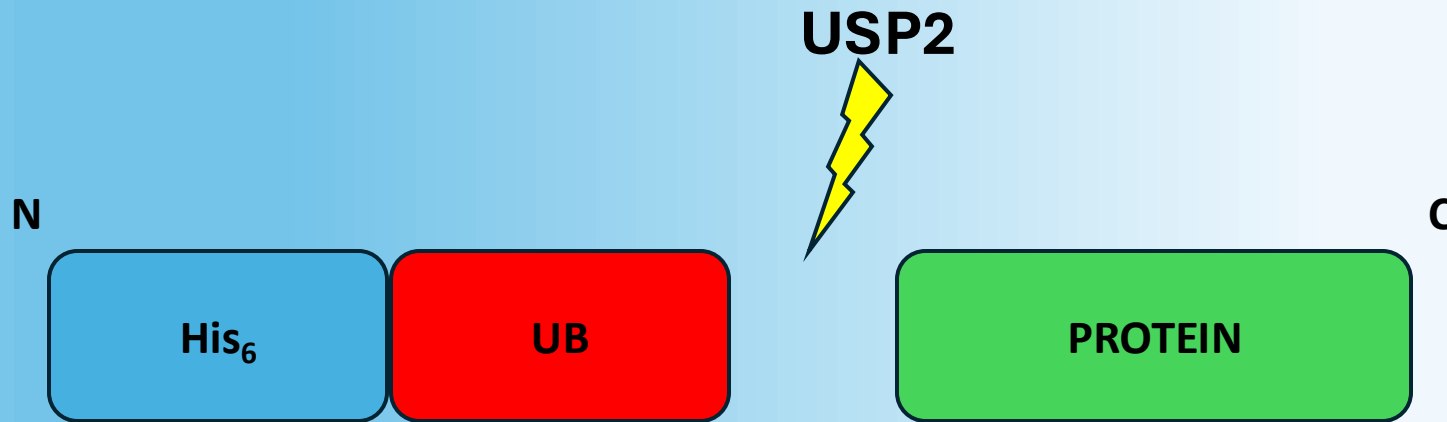


Histidine

His-Ubiquitin tag removal



The ubiquitin portion of the tag allows easy removal of the entire fusion partner



The His-Ubiquitin tag is removed through digestion with USP2

The resulting protein is now free of exogenous portions

Further processing

The recombinant protein can be purified further to eliminate remaining contaminants

To this end, additional chromatography techniques are employed, such as ion exchange and size exclusion chromatography





Thank you for your attention!

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